An rbrothers tutorial

Jan Irvahn and Vladimir N. Minin
Department of Statistics, University of Washington Seattle, WA, 98195, USA

This is a tutorial demonstrating the usage of the R package `rbrothers` that provides an easy R interface to java program, `DualBrothers` [2], and adds a number of auxiliary functions to make `DualBrothers` more user friendly. To get started, install the `rbrothers` R package from R-forge.

```
> install.packages("rbrothers", repos="http://R-Forge.R-project.org")
```

The rJava package and a couple of other packages will need to be installed.

Start R and load the rbrothers library.

```
> library(rbrothers)
```

We walk through the rbrothers workflow using an HIV circulating recombinant form KAL153 from Kaliningrad, Russia [1], aligned with genomic sequences of HIV-1 subtypes A, B, and F. This is one of the examples from the original `DualBrothers` paper [2]. First, copy the KAL153.phy file from the rbrothers package to your current working directory.

```
> my.align = read.dna(file = system.file("extdata/KAL153/KAL153.phy",
     package = "rbrothers"))
> write.dna(my.align, "KAL153.phy")
```

Now you can run DualBrothers with a single command. The 'seed' argument is any integer used to seed the package's pseudo-random number generator. The 'alignment' argument is the name of the input file (excluding the '.phy' extension). The 'format' argument tells rbrothers the format of the input file, either 'interleaved' or 'fasta'. The 'par_lambda' argument sets the prior mean number of substitution process change-points. The 'top_lambda' argument sets the prior mean number of topology breakpoints. You can find more information about the details of the DualBrothers program at `http://faculty.biomath.ucla.edu/msuchard/htdocs/DualBrothers/`.

```
> db <- dualbrothers(seed = 123, alignment = "KAL153",
     format = "interleaved", par_lambda = 5, top_lambda = 0.693)
```

When your alignment has more than six tips `rbrothers` constructs a set of candidate trees. You need to provide a value for the 'window.size' argument in this situation as mentioned in the help file: `help(dualbrothers)`. If the set of candidate trees is too small you have the option to bootstrap additional candidate trees via the 'boot' argument.

If you already have the output files of a DualBrothers run you can read the information in directly.

```
> db <- readdb("KAL153")
```

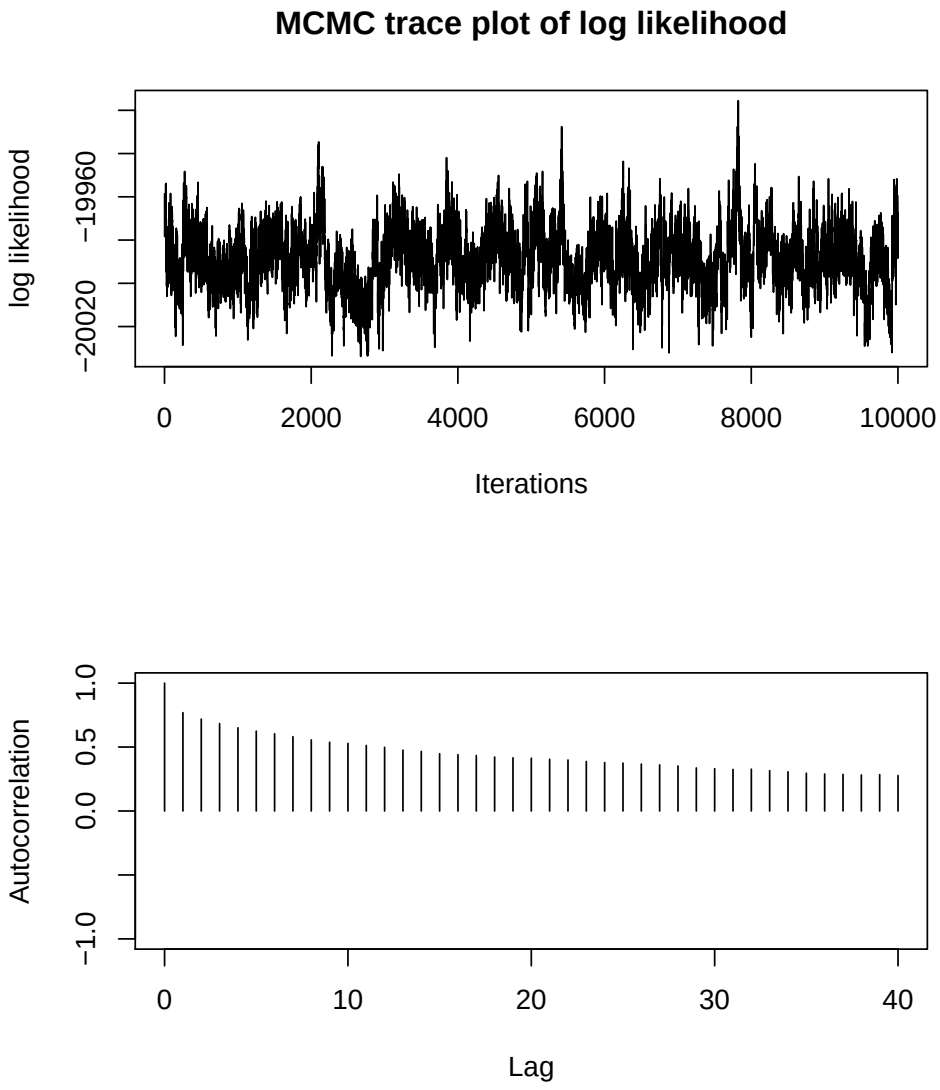

Figure 1: The top plot shows the trace plot of the log likelihood of the MCMC chain. The bottom plot shows the autocorrelation of the log likelihood of the MCMC chain.

It is a good idea to check the convergence of the MCMC chain; if the chain has not converged it should be rerun with a larger number of MCMC iterations. The 'length' argument of the dualbrothers command allows the user to specify how long the MCMC chain should be. A trace plot of the log likelihoods of the MCMC chain and an autocorrelation plot can be created with a single command.

```
> plotmcmc(db)
```

The trace plot indicates that the MCMC chain reached stationarity; the autocorrelation plot shows significant autocorrelation when subsampling the chain every 200 iterations. When increasing the length of a chain we suggest increasing the subsampling parameter, 'subsam-

ple', so as to avoid saving large amounts of output. The 'burnin' parameter controls the number of iterations discarded at the beginning of the MCMC run.

Summary plots are easily created in `rbrothers`. The plot command produces two graphs on top of each other. The bottom plot is the posterior probability of a breakpoint at each nucleotide position. The top plot is the posterior probability of each tree at each nucleotide position. The trees are color coded; the user can provide a list of their preferred colors via the 'color' argument. The help file for plot.db explains the arguments of the plot command: `help(plot.db)`.

```
> plot(db)
```

A separate command, `plottree.db`, can be used to see the most probable trees. This function uses ape's `plot.phylo` command so the type argument (controlling the style to display the phylogenetic tree) can be set to "phylogram", "cladogram", "fan", "unrooted", or "radial".

```
> plottree.db(db, type = "phylogram")
```

The threshold argument allows you to only plot trees that have a posterior probability greater than the threshold provided at some point along the alignment. The 'seetrees' argument allows the user to replace the posterior probability of a breakpoint plot with the trees associated with the posterior tree probability plot.

```
> plot(db, seetrees = TRUE, threshold = 0.5, type = "cladogram")
```

The average number of breakpoints, the posterior probability of at least one breakpoint and DualBrothers parameters associated with the run can be easily accessed:

```
> summary(db)

DualBrothers output for KAL153
4 sequences of length 8588

MCMC settings:
length of the MCMC chain: 2100000
burn-in length: 100000
subsample frequency: 200

Prior parameters:
prior mean number of substitution process change points: 5
prior mean number of topology change points: 0.693

average number of breakpoints in the posterior: 2.1
The posterior probability of at least one breakpoint was 1
(> 0.999  required for a Bayes factor > 1000).
3 trees considered
```

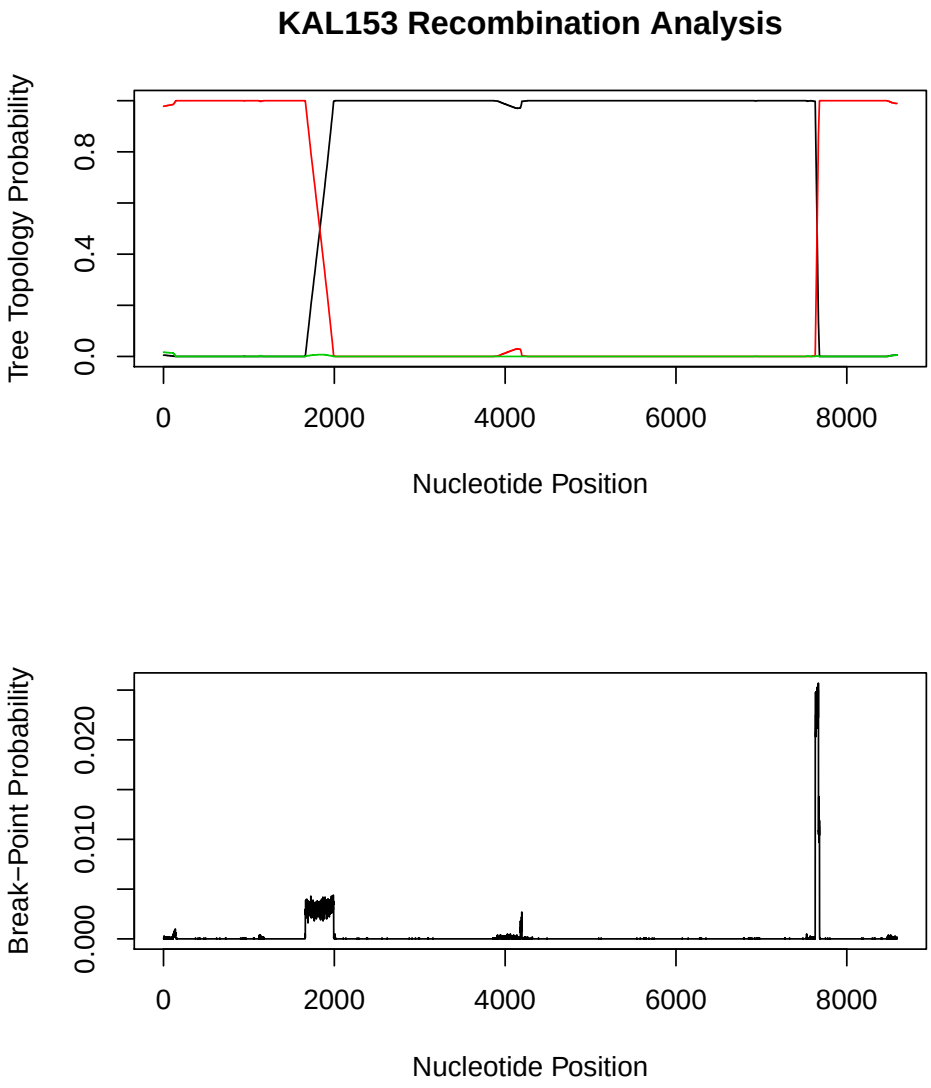

Figure 2: The top plot shows site specific posterior probabilities for the most probable phylogenetic tree topologies. The bottom plot shows the site specific posterior probability of a breakpoint.

You can calculate a 95% Bayesian credible interval for a breakpoint if you can provide a nucleotide position interval that contains exactly one breakpoint, for example, $(1000, 3000)$.

```
> breakpointCI(db, 1000, 3000)

The 95% credible interval for a single break point between nucleotide
number 1000 and nucleotide number 3000 is:
 2.5% 97.5%
 1666  1985
```

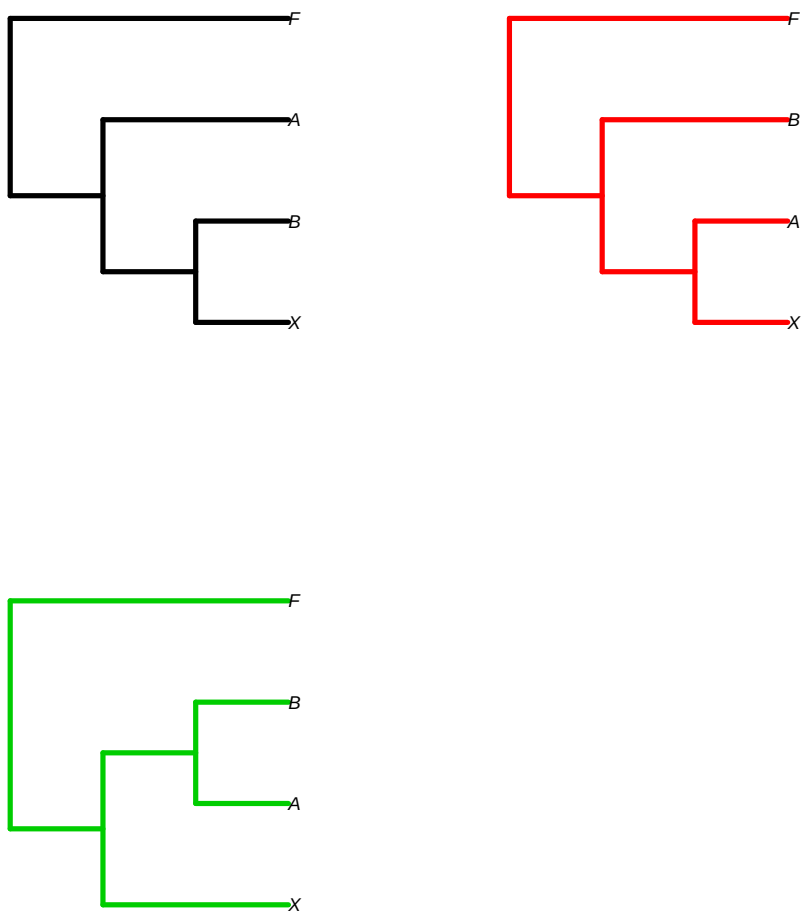

Figure 3: These are the most probable phylogenetic tree topologies.

References

[1] Liitsola,K. *et al.* (1998) HIV-1 genetic subtype A/B recombinant strain causing an explosive epidemic in injecting drug users in Kaliningrad. *AIDS,* **12**, 1907-1919.

[2] Minin,V. *et al.* (2005) Dual multiple change-point model leads to more accurate recombination detection. *Bioinformatics*, **21**, 3034-3042.

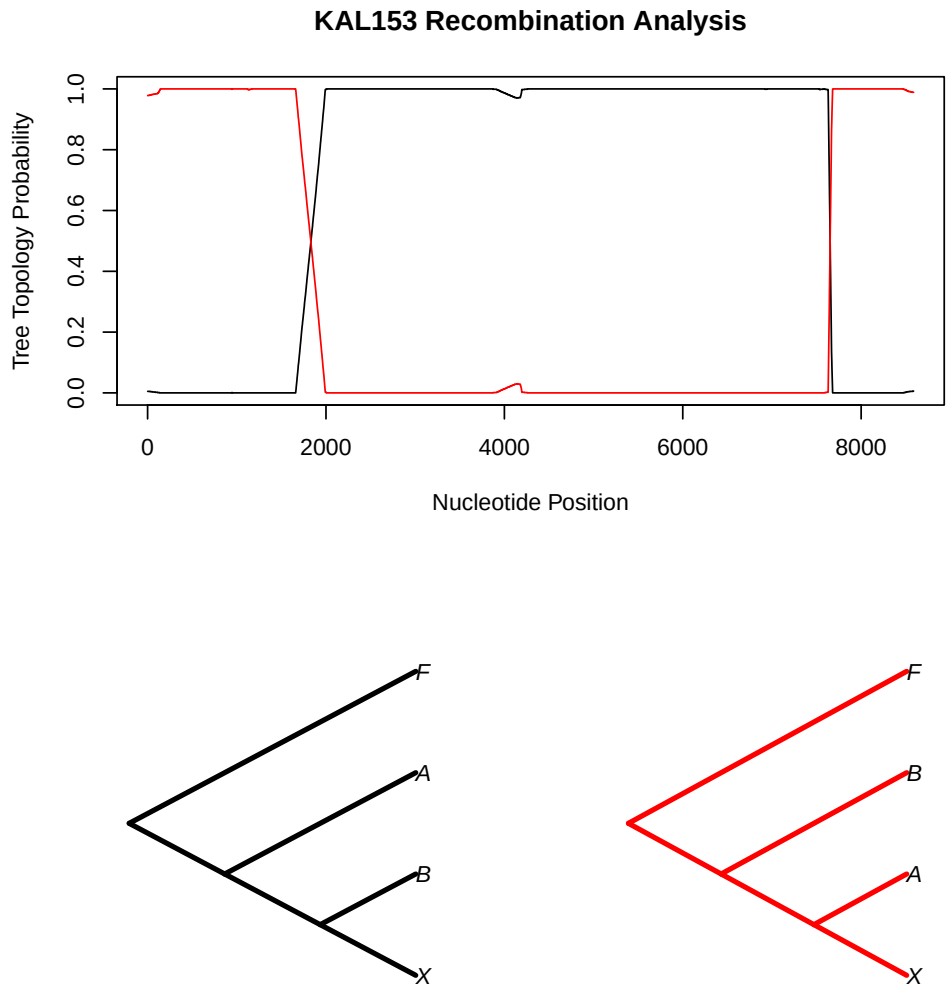

Figure 4: The top plot shows site specific posterior probabilities for the two phylogenetic trees whose posterior probability exceeded the specified threshold, 0.5. The bottom plot shows the two trees.